



Dictyanthus reflexiflorus (Apocynaceae: Asclepiadoideae) a new species from Mexico

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Abstract

A new species of *Dictyanthus* from the state of Michoacán, Mexico, is described and illustrated *Dictyanthus reflexiflorus* sp. nov. It grows in tropical deciduous forests located in the municipality of Tacámbaro. A phylogenetic analysis based on molecular data (plastid regions *trnL-trnF* and *rps 16*) was conducted to explore the relationship of the new species. Results place the new species in *Dictyanthus*. The morphological characters that support this relationship are the presence of uncinat hairs and a reticulate corolla. The species has unique characteristics within the genus, such as a reflexed corolla and the tube 10-lobed closely pressed to the side of the gynostegium. A key to distinguish *Dictyanthus reflexiflorus* from congeners in Mexico is included.

Key words: Asclepiadoideae, endemic, Gonolobinae, Michoacán, phylogeny

Introduction

Dictyanthus Decaisne (1844: 604) belongs to the subtribe Gonolobinae (Apocynaceae, Asclepiadoideae) and contains 18 species distributed in Central America, from northern Mexico to Nicaragua, with 87% of its species endemic to Mexico (Stevens 2001; Stevens 2009; Endress *et al.* 2019; González-Martínez *et al.* 2019; Alvarado-Cárdenas *et al.* 2020; Alvarado-Cárdenas *et al.* 2022). The genus was considered a subgenus of *Matelea* Aublet (1775: 277) due to the similar flowers and a mixed indumentum of long straight, short, and glandular trichomes (Woodson 1941; Stevens 1988; Krings *et al.* 2008; Endress *et al.* 2014).

Recent phylogenetic analyses recovered *Matelea s.l.* as paraphyletic, favoring the reinstatement of the *Dictyanthus* clade as a genus (McDonnell *et al.* 2018; González-Martínez *et al.* 2019). *Dictyanthus* can be distinguished from the rest of the subtribe by a palmately 5-lobed gynostegial corona, confluent with the corolla lobes, uncinat trichomes on the vegetative parts, and follicles with a ridge at the base and prickly projections (González-Martínez 2019; Stevens 1988; Stevens 2000; Stevens 2001; Stevens 2009). The corolla in *Dictyanthus* is shallowly or deeply campanulate to urceolate sacciform (Stevens 1988; Stevens 2009).

During fieldwork in the state of Michoacán, Mexico, we collected specimens that could not be assigned to any known species of *Dictyanthus*. We conducted phylogenetic analyses to determine the relationships of this new species and we describe it as *Dictyanthus reflexiflorus*. We also provide a key to the species of *Dictyanthus* found in Mexico.

Materials and methods

Extraction, amplification, and sequencing of DNA

Total genomic DNA of *Dictyanthus reflexiflorus* was extracted from leaves using the CTAB method (Martínez-González *et al.* 2017). The DNA was quantified with a Nanodrop 2000c (Thermo, USA). We prepared dilutions at 20 ng. Three molecular markers were used: the *rps* 16 intron, was amplified using the primers *rpsF* and *rpsR* (Oxelman *et al.* 1997); the *trnL* intron, and the *trnL-trnF* spacer, both markers amplified with the primers C, D, E, F (Taberlet *et al.* 1991). The PCR reaction mixture was prepared in a final volume of 15 µL containing buffer of the enzyme 1x Taq DNA polymerase, 0.8mM deoxynucleoside triphosphate (0.2mM of each one), 100 ng DNA, 20 pmol of each primer and 2 units of GoTaq DNA (Promega, USA). The amplification of each region was undertaken through an initial denaturing cycle at 94°C for four minutes followed by thirty-five cycles of denaturing at 94°C for forty-five seconds, annealing for one minute at the specific temperature of each gene, and extension at 72°C for five minutes. All PCR reactions were conducted in a Peltier Thermal Cycler PTC-200 (BIORAD, México). The amplifications were verified through electrophoresis in a 1.5% agarose gel prepared with 1× TAE buffer (Tris-Acetate-EDTA) and run at 87V cm⁻³ for one hour. The gel was stained with GelRed (Biotium, USA), and the bands were visualized in an Infinity 3000 transilluminator (Vilber Lourmat, Germany). The amplified products were purified with the ExoSAP Purification kit (Affymetrix, USA), following the manufacturer's instructions. These products were sequenced using an Applied Biosystems model 3730XL (Applied BioSystems, USA) at the Biology Institute of the Universidad Nacional Autónoma de México.

Sequence assembly

The forward and reverse sequences of each of the three regions were edited and assembled using BioEdit version 7.0.5 (Hall 1999) to generate a consensus sequence. The sequences were aligned separately using MAFFT v. 7 (Katoh & Standley 2013) with the FFT-NS-i iterative refinement method, and the rest of the parameters selected by default. Subsequently, the alignments were manually edited using PhyDE v. 0.9971 (Müller *et al.* 2010).

Phylogenetic analysis

We downloaded sequences which had been used in previous phylogenetic studies of Gonolobinae (Liede-Schumann *et al.* 2005; Krings *et al.* 2008; Mangelsdorff *et al.* 2016). Taxa were chosen from the species with data from the molecular markers that we sequenced. The complete list of GenBank accession numbers for the species included is presented in Appendix 1.

For the substitution model of each marker, we used Model Finder (Kalyaanamoorthy *et al.* 2017) employing default parameters and chosen under the Akaike information criteria (AIC). Maximum Likelihood Analysis (ML) was performed in IQ-TREE web server (Trifinopoulos *et al.* 2016) with ultrafast bootstrapping (Hoang *et al.* 2018) of 1000 replicates and the rest of parameters selected by default. The consensus tree and bootstrap percentages (BS) were viewed and edited in Fig Tree v. 1.4.3 (Rambaut 2016). Bayesian Inference (BI) was performed using Mr. Bayes v. 3.2.6 (Ronquist *et al.* 2012) with two independent runs and four Markov chains Monte Carlo (MCMC) each. Each chain ran for 10 million generations and was sampled every 1,000 generations. The convergence of the chains was checked using Tracer v. 1.7.1 (Rambaut *et al.* 2018), and according to the performance of the chains, 25% of the trees were eliminated as burn-in. The remaining trees were used to build a consensus tree, which was viewed using FigTree v. 1.4.3 (Rambaut 2016). The aligned sequences and resulting trees are available in Dryad (Martínez-Ambriz *et al.* 2023).

Morphological observations

Material of the new species was collected during two trips to the state of Michoacán (municipality of Tacámbaro) in September 2015 and September 2016. Specimens were deposited in the herbaria FCME, IEB, and MEXU (acronyms according to Thiers, 2021). Additionally, flowers were fixed in 70% alcohol for detailed observations with a dissecting microscope. This material was compared with all known species of *Dictyanthus* (Stevens 1988; Stevens 1999; Stevens 2000; González-Martínez *et al.* 2019; Alvarado-Cárdenas *et al.* 2022), from specimens in the herbaria ENCB, FCME,

MEXU, and XAL. Those species found in Mexico were included in a key with the new species. IUCN criteria (IUCN 2022) were used to assess the conservation status of the new species.

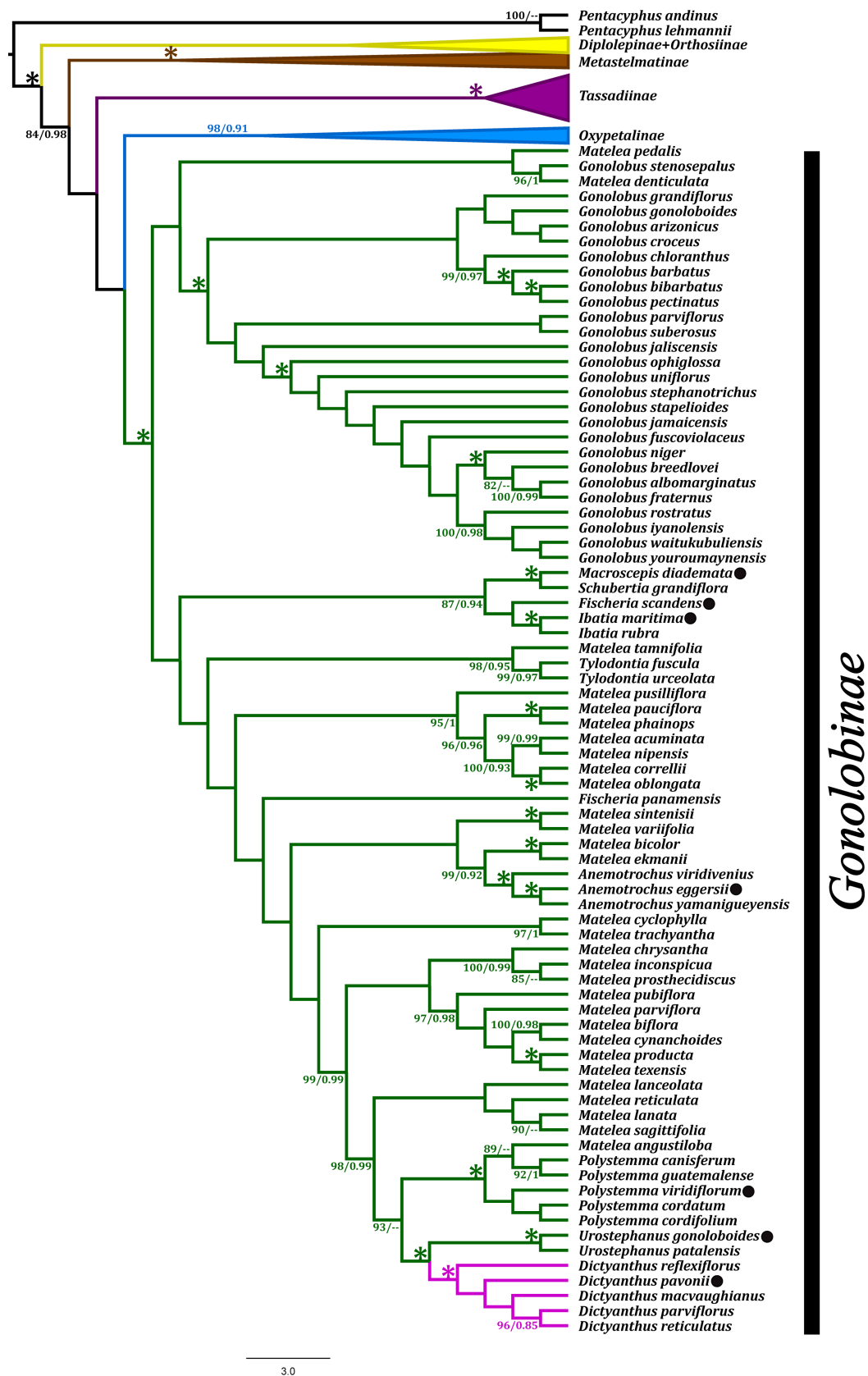


FIGURE 1. Tree derived from Bayesian Inference, showing the position of *Dictyanthus reflexiflorus* in Gonolobinae. Numbers on branches indicate Maximum Likelihood bootstrap support (BS) / Posterior Probability (PP). Positions where BS = 100 and PP = 1.0 are indicated by asterisks. Only BS \geq 80 or PP \geq 0.8 are indicated. Type species included in the analysis are indicated by a circle. To save space the clades of *Diplolepineae+Orthosiniinae*, *Metastelmatinae*, *Oxypetalinae* and *Tassadiinae* were collapsed.

Results

A summary of the data from the aligned sequences is presented in Table 1. The marker *rps16* was the most variable (16.4% variable sites), followed by *trnL-trnF* (9.3%).

TABLE 1. Summary of statistics for the analyzed markers used to run ML and BI analyses.

Region	<i>rps16</i>	<i>trnL-trnF</i>
No. of taxa	172	172
Aligned length (bp)	947	1067
Conserved characters	662	829
Variable characters	155	99
Substitution model	TVM+F+I+G4	TVM+F+G4

ML and BI analyses recovered Gonolobinae as monophyletic (BS = 100; PP = 1), which is consistent with previous studies. Both BI and ML analyses, recovered the four species of *Dictyanthus* as monophyletic. The new species, *Dictyanthus reflexiflorus*, is sister (BS = 100, PP = 1; Fig. 1) to these four congeners included here: *D. macvaughianus* (Stevens) Stevens (2000: 243), *D. parviflorus* Hemsley (1882: 329), *D. pavonii* Decaisne (1844: 605) (the type of *Dictyanthus*) and *D. reticulatus* (Turcz.) Bentham & Hooker f. ex Hemsley (1882: 329). The position of the new species was the same in the trees from BI and ML analyses. *Dictyanthus* was recovered as sister to a clade (BS = 100; PP = 1), composed of *Urostephanus gonoloboides* Robinson & Greenman (1895: 159–160) (the type of *Urostephanus* Robinson & Greenman (1895: 159)) and *U. patalensis* (Donn. Sm.) González-Martínez, Lozada-Pérez & Alvarado (2024: 13). Both clades are deeply nested within *Matalea*.

Taxonomy

Dictyanthus reflexiflorus Lozada-Pérez, E.B. Cortez & Martínez-Ambriz, *sp.nov.* (Figs. 2, 3).

Type:—MEXICO. Michoacán: Municipio Tacámbaro. Camino de Chipícuaro a Las Ánimas, 19°09'07"N, 101°23'00"W, 1170 m, 12 Sept 2016, L. Lozada-Pérez, E. Martínez-Ambriz & L. Pérez-García 3835 (holotype: FCME!; isotypes: IEB!, MEXU!).

Dictyanthus reflexiflorus shares a climbing habit with *D. altatensis* (Brandege) Stevens (2000: 243) but differs from it by the reflexed corolla 10-lobed at base and inflorescence with peduncles 3–4 mm long (vs. corolla rotate, without lobes at the base and inflorescence with peduncles 4–8 cm long).

Herbaceous climber to 2.5 m long, with a woody base, latex clear. *Stems* and branches cylindrical, violet to purple, slightly corky, lignified at the base and glabrescent with age; young stems with evenly distributed mixed indumentum composed for multicellular and unicellular trichomes, multicellular ones can be hooked or straight, 0.75–1.25 mm long, yellowish, unicellular trichomes are glandular or eglandular, less than 0.1 mm long, grayish. *Leaves* opposite, without stipules, petioles 1–3 cm long, with indumentum similar to the stem; lamina membranaceous, 2.0–7.0 × 1.0–3.5 cm, ovate, 5 or 6 secondary veins, adaxial surface sparsely pubescent with scattered uncinata hairs, slightly denser on the midvein near the base, abaxial surface densely puberulent with short uncinata trichomes and scattered glandular trichomes, veins with scattered long uncinata trichomes; base cordate, apex acuminate, base cordate; colleters 4–6, at the adaxial junction of petiole and lamina. *Inflorescence* extra-axillary, one per node, umbelliform, with 3–5 flowers; peduncle 3–4 mm long; bracts 3.0–4.0 × 0.3–0.5 mm, narrowly lanceolate; pedicels 9.0–11.0 mm long. *Flowers* pentamerous. *Calix* green, lanceolate, 4.5–5.0 × 1.5–2.0 mm, pubescent with widely scattered uncinata trichomes on the outside and shorter trichomes on the margin; colleters 1 per sinus. *Corolla* reflexed, pale to greenish yellow with dark reticulation, adaxially densely hirsute, abaxially densely hirsute with short uncinata trichomes and scattered glandular trichomes, tube 3.0–3.5 mm long, formed by erect slightly 10-lobed closely pressed to the side of the gynostegium and reaching to just below the corona, lobes ovate-lanceolate, 4.5–5.0 × 4.0–4.3 mm; gynostegial corona black, 5-lobed, lobes alternating with anthers, ca. 1.5 × 0.5 mm, each lobe 2-keeled, each keel parallel to

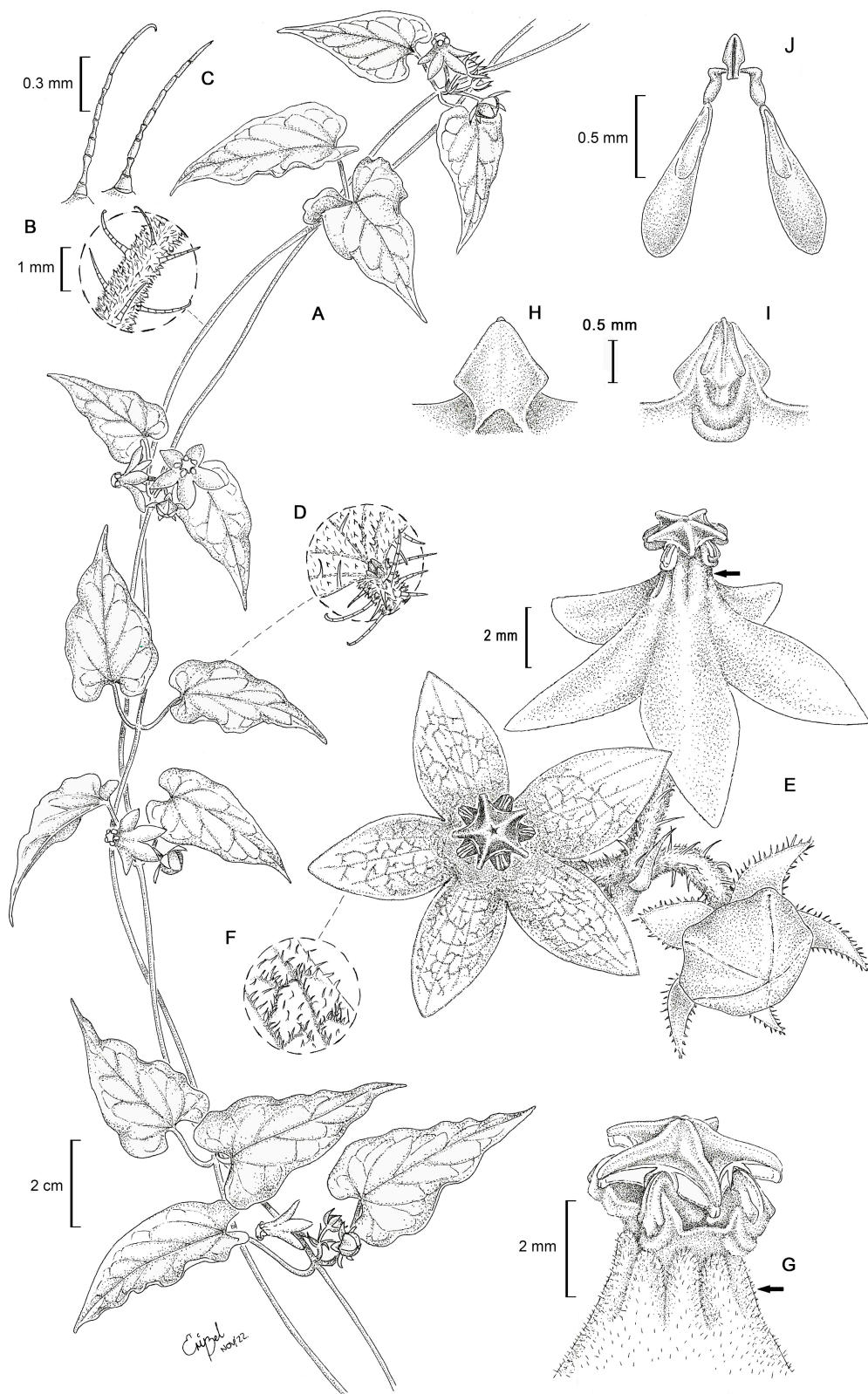


FIGURE 2. *Dictyanthus reflexiflorus*. A. Plant with inflorescences. B. Close-up of branch showing mixed indumentum. C. Close-up of thichomes. D. Colleters at base of leaf-blade. E. Inflorescence. F. Close-up of the indument of the abaxial surface of the corolla. G. Gynostegium with style-head. H. Gynostegial corona lobe in posterior view. I. Gynostegial corona lobe in frontal view. J. Pollinarium. Arrows showing the corolla lobes. Illustrations by Ericka B. Cortez, based on Lozada-Pérez *et al.* 3835.

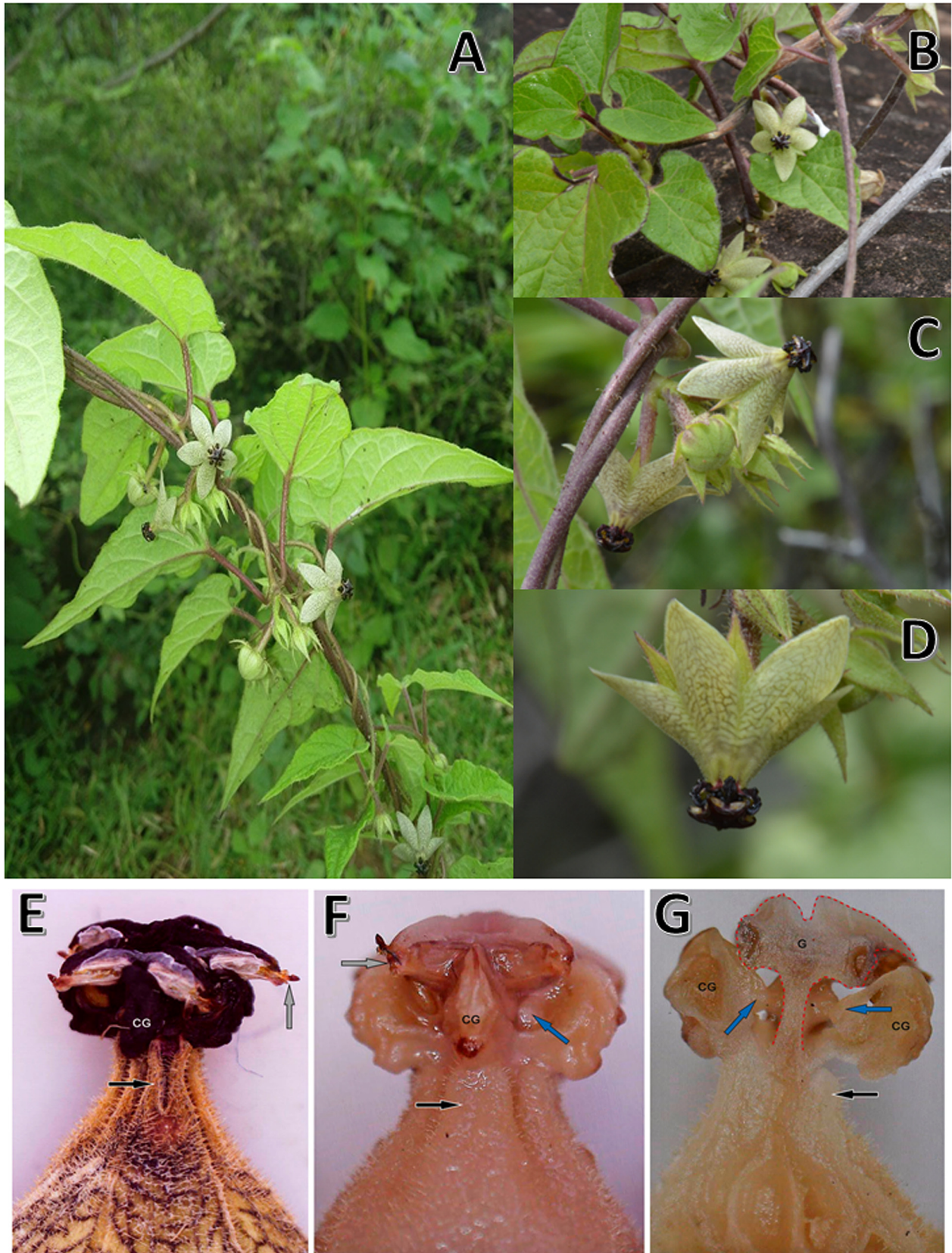


FIGURE 3. *Dictyanthus reflexiflorus*. A-C. Plant and inflorescences with mature flowers. D. Side view of corolla and gynostegium. E. Dried flower. F. Flower fixed in alcohol. G. Vertical section of corona and gynostegium. (GC: Gynostegial corona, G: Style head, black arrows = tube lobes, gray arrows = corpuscle and blue arrows = keels). Photos by María de la Luz Perez-Garcia and Lucio Lozada-Pérez.

the adjacent keel; gynostegium, ca. 4 mm diam., deeply pentagonal, convex above; pollinarium 1.1–1.4 mm long, corpusculum dark brown, slightly sagittate, ca. 0.2×0.07 mm, caudicles 0.17–0.2 mm long, pollinia elliptic, 0.8–1.0 mm long, apically excavated. *Follicles* unknown.

Distribution and Habitat:—*Dictyanthus reflexiflorus* is distributed in a small area of central Michoacán (Fig. 4), in the Balsas Basin province (Morrone *et al.* 2017), in tropical deciduous forest, from 1170–1185 m.

Phenology:—Flowering has been recorded in September.

Etymology:—The epithet *reflexiflorus* refers to the reflexed corolla, a distinctive feature that is unique in *Dictyanthus*.

IUCN conservation assessment:—Information about *Dictyanthus reflexiflorus* is scarce, with only one population known, although the surrounding area was explored, ca. 1 km². For this reason, we suggest that it be assigned the category of Data Deficient (DD) (IUCN 2022) until a thorough evaluation is conducted in the adjacent hills and ravines. The type locality is not part of a natural protected area, and we have not carried out an extensive vegetation survey of the area.

Taxonomic notes:—*Dictyanthus reflexiflorus* is very distinctive due to its reflexed corolla, with a tube 10-lobed closely pressed to the side of the gynostegium (Fig. 3 E, G). All other species have a rotate or campanulate corolla, which is smooth towards its base inside (Stevens 1988, 1999, 2000; González-Martínez *et al.* 2019; Alvarado-Cárdenas *et al.* 2022). Similarly, the very reduced size of the gynostegial corona, which is limited to the base of the limb, is different from that in other species where the corona is almost completely adnate to the corolla. This unusual structure is contrasted by the star-shaped style-head with an unusually dark.

The new species is endemic to the state of Michoacán and known only from two collections from the same locality. The climbing habit is shared with *D. altatensis*, but in this species the corolla is rotate, and the tube is not 10-lobed at the base.

Additional specimens examined:—MEXICO. Michoacán: Tacámbaro, 2.19 km camino de Chipícuaro a Las Ánimas, 19°09'06"N, 101°23'00"W, 1185 m, 3 Sept 2015, L. Pérez G. *et al.* 163 (IEB).

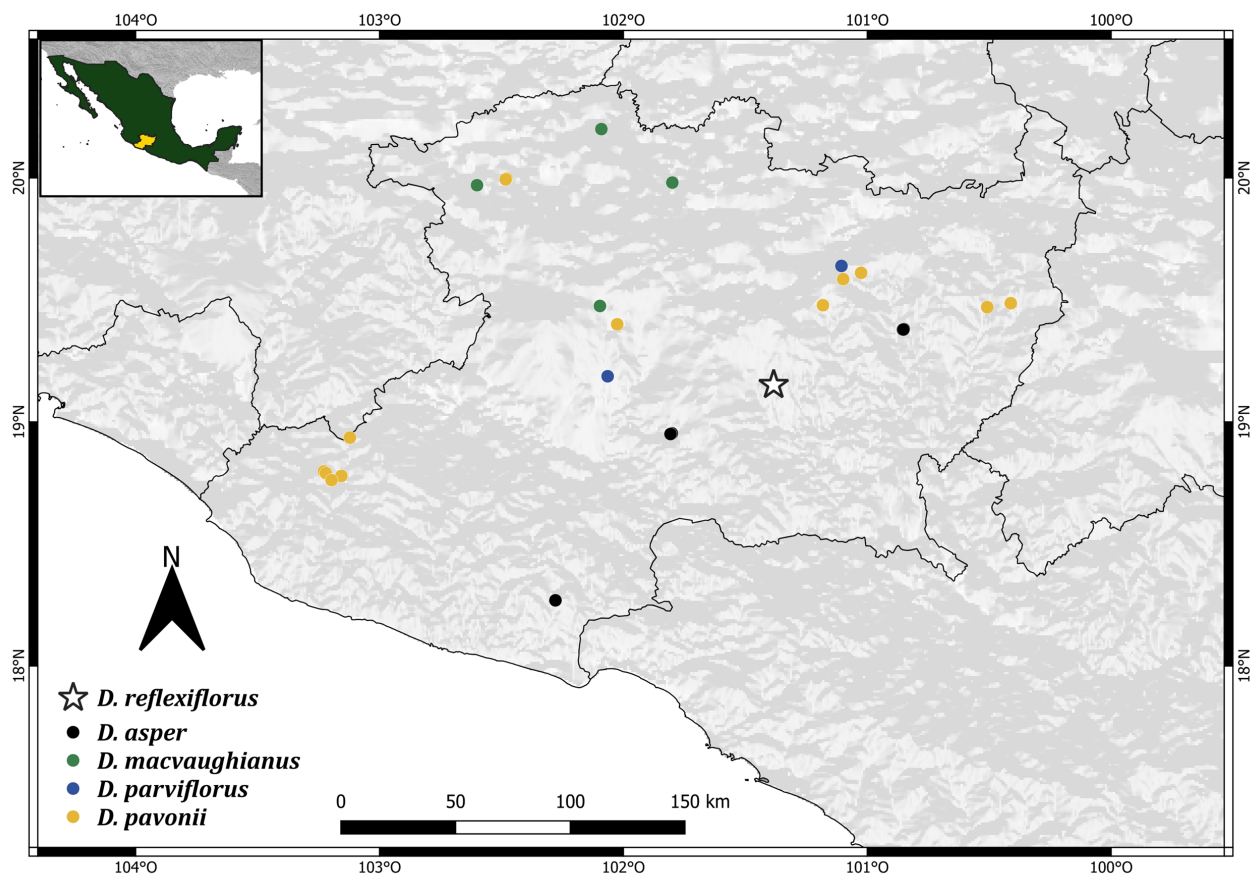


FIGURE 4. Distribution of *Dictyanthus reflexiflorus* and their congeners in Michoacan, Mexico.

Discussion

Our result show that the Gonolobinae is monophyletic, agreeing with previous studies (Liede-Schumann *et al.* 2005; Krings *et al.* 2008; Mangelsdorff *et al.* 2016; McDonnell *et al.* 2018). Within the Gonolobinae, several other genera were also recovered and strongly supported (Mangelsdorff *et al.* 2016; McDonnell *et al.* 2018; González-Martínez *et al.* 2024), including the relationship between the two subclades of *Gonolobus* (McDonnell *et al.* 2018).

The four species of *Dictyanthus* included in our analysis formed a strongly supported clade together with our new species. Although *D. reflexiflorus* not show some diagnostic taxonomic characters from *Dictyanthus*, like the palmately 5-lobed gynostegial corona (Gonzalez-Martínez 2019), shared the uncinete trichomes on the vegetative parts and the reticulation of the inner surface corolla. These morphological evidence with the molecular and distribution data allow us to locate these species like a new member of *Dictyanthus*. The uncommon floral morphology in the new species first made us think in a new genus of Gonolobinae. However, the available data do not support this hypothesis and include this species like a new member of *Matelea s.l.* seems impractical to us. The latest classification of Apocynaceae (Endress *et al.* 2014) did not recognize *Dictyanthus* as an independent genus. But the most recent phylogenetic analysis shows *Matelea s.l.* like a paraphyletic genus with a lack of discrete morphological variation (Morillo 2015; McDonnell *et al.* 2018; González-Martínez 2019) and support the monophyly of *Dictyanthus* (Krings *et al.* 2008; McDonnell *et al.* 2018; González-Martínez 2019; González-Martínez *et al.* 2024). Here, we follow the proposal of *Dictyanthus* as an independent genus, which allow a better understanding of the group. Our results highlight the high diversity in *Dictyanthus* and distinguishes Gonolobinae as a source of botanical novelties.

Identification key for *Dictyanthus* in Mexico (modified from González-Martínez 2019)

1. Corolla tube adaxially with parallel vertical lines, occasionally with perpendicular lines2
1. Corolla tube adaxially with concentric lines or without perpendicular lines or lacking a distinctive pattern7
2. Pedicels less than 6 mm long; corolla lobes less than 7 mm long3
2. Pedicels more than 6 mm long; corolla lobes more than 7 mm long6
3. Inflorescence with 2 flowers; corona lobes oblong or linear-spathulate, more than half the length of the corolla tube4
3. Inflorescence with more than 2 flowers; corona lobes rhombic or triangular, less than half the length of the corolla tube5
4. Leaf (3.1–)3.6–5.6 × 2–4.3(–5) cm; petioles 1.6–4.6 cm; corolla green; corona lobes oblong with rounded or bilobed apex
.....*D. stevensii*
4. Leaf 10.7–13 × 7.1–8.8 cm; petioles 4.7–8.1 cm; corolla white-pink; corona lobes linear-spathulate with elliptical apex
.....*D. lautus*
5. Plant a climber; pedicels 1.5–3.5 mm long; calyx lobes 3–5 mm long; corolla brownish-greenish or whitish; corona-lobes rhombic
.....*D. sepicola*
5. Plant erect; pedicels 4–5 mm long; calyx lobes 5–9 mm long; corolla grayish-white; corona lobes triangular*D. tuberosus*
6. Corolla whitish-yellow; peduncle and pedicels with generally uncinete trichomes; corona with a thin band of short trichomes around and inside the lobes, lobes 6–10 mm long*D. pavonii*
6. Corolla yellowish-green; peduncle and pedicels with straight trichomes; corona glabrous around and inside lobes, lobes 11–25 mm long
.....*D. macvaughianus*
7. Corolla glabrous or almost so within; corona lobes basally connate forming a disc8
7. Corolla densely pubescent within, at least on the limb; corona lobes sometimes connate, but without forming a disc9
8. Sepals (3.5–)5–10 × (1–)2.4 mm; corolla rotate, white or green with reticulated venation; lobes 3.5–8 mm long; corona lobes vesicular lobes
.....*D. asper*
8. Sepals 10–17 × 4–6.5 mm; corolla campanulate, pale yellow with concentric venation, lobes 8–13 mm long; corona lobes subulate
.....*D. hamatus*
9. Corona lobes oblong with spherical and roughened apex10
9. Corona lobes not as above12
10. Sepals more than 5 mm long; corona lobes as long as corolla tube11
10. Sepals less than 5 mm long; corona lobes longer than corolla tube*D. suffruticosus*
11. Corolla lobes revolute, limb and lobes slightly reflexed; grey-purple corolla in fresh with reticulated venation
.....*D. yucatanensis*
11. Corolla lobes not revolute, limb and lobes patent or slightly ascending; yellowish-green corolla when fresh without obvious venation
.....*D. aeneus*
12. Corona lobes triangular or keeled, less than 2 mm long13
12. Corona lobes linear or linear-spathulate, more than 4.5 mm long15
13. Plant erect; leaf 1.3–3.4 cm long; pedicel 3–5 mm long*D. parviflorus*
13. Plant a climber; leaf 2–8.7 cm long; pedicel (6–)9–28 mm long14
14. Corolla rotate, adaxially without lobes at the base; inflorescences with peduncles 4–6 cm long*D. altatensis*
14. Corolla reflexed, adaxially 10-lobed at base; inflorescences with peduncles 3–4 mm long*D. reflexiflorus*
15. Limb of the corolla with concentric venation16
15. Limb of the corolla with irregular concentric-reticulate venation17

16. Corolla tube with concentric venation; corona lobes linear lobes..... *D. tigrinus*
 16. Corolla tube with parallel vertical venation; corona lobes linear-spatulate*D. eximus*
 17. Corona lobes 5–8 mm long; style-head concave above*D. reticulatus*
 17. Corona lobes 8–11 mm long; style-head apiculate above*D. ceratopetalus*

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APPENDIX 1. Sequences from GenBank used in this study, including taxa and GenBank accession numbers. Accession numbers that include the *trnL* intron and the *trnL-trnF* spacer are indicated with an asterisk.

Species	<i>rps16</i>	<i>trnL</i> intron	<i>trnL-trnF</i> spacer
<i>Pentacyphinae</i>			
<i>Pentacyphus andinus</i>	AJ699335	AJ492151	AJ492150
<i>Pentacyphus lehmannii</i>	AJ704928	AJ290888	AJ290889
<i>Anemotrochus eggersii</i>	HF547187	HF547131*	
<i>Diplolepiniae</i>			
<i>Diplolepis boerhaviifolia</i>	AJ699331	AJ428608	AJ428607
<i>Diplolepis geminiflora</i>	AJ699332	AJ410183	AJ410182
<i>Diplolepis hieronymi</i>	AJ699333	AJ410213	AJ410212
<i>Diplolepis menziesii</i>	AJ699276	AJ699275	AJ699273
<i>Diplolepis nummulariifolia</i>	AJ699334	AJ290851	AJ290852
<i>Orthosüinae</i>			
<i>Jobinia formosa</i>	AJ699346	AJ428641	AJ428640
<i>Jobinia hatschbachii</i>	HE611842	HE611802	HE611763
<i>Jobinia umbellata</i>	AJ704319	AJ704318	AJ704316
<i>Monsanima morrenioides</i>	AJ699348	AJ428686	AJ428685
<i>Orthosia ellemannii</i>	AJ699350	HE611812	HE611773
<i>Orthosia kunthii</i>	HE611857	HE611813	HE611774
<i>Orthosia scoparia</i>	HE611862	AY163703*	
<i>Scyphostelma beckii</i>	AJ704304	AJ704306	AJ704307
<i>Scyphostelma bifidum</i>	HE611873	HE611829	HE611790
<i>Scyphostelma ecuadorensis</i>	HE611874	HE611830	HE611791
<i>Scyphostelma harlingii</i>	AJ704311	AJ704310	AJ704308
<i>Scyphostelma microphyllum</i>	AJ699347	HE611831	HE611792
<i>Metastelmatinae</i>			
<i>Barjonia chloraeifolia</i>	AJ704463	AY163667*	
<i>Barjonia glazioui</i>	JN574479	JN574618*	
<i>Barjonia laxa</i>	JN574481	JN574620*	
<i>Blepharodon ampliflorum</i>	JN574482	JN574620*	
<i>Blepharodon bicuspidatum</i>	JN574483	JN574621*	
<i>Blepharodon glaucescens</i>	AJ699292	AJ699291	AJ699289
<i>Blepharodon grandiflorum</i>	AJ699337	AJ290838	AJ290837
<i>Blepharodon lineare</i>	AJ704466	AY163668*	
<i>Blepharodon mucronatum</i>	AJ699338	AJ290839	AJ290840
<i>Blepharodon pictum</i>	AJ704467	AY163669*	
<i>Ditassa banksii</i>	AJ704473	AY163674*	
<i>Ditassa burchellii</i>	AJ699293	AJ699295	AJ699296

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APPENDIX 1. (Continued)

Species	rps16	trnL intron	trnL-trnF spacer
<i>Ditassa decussata</i>	AJ704217	AJ704219	AJ704220
<i>Ditassa ditassoides</i>	AJ704476	AY163678*	
<i>Ditassa endoleuca</i>	HE611840	HE611801	HE611762
<i>Ditassa hispida</i>	AJ704477	AJ704480	AJ704478
<i>Ditassa micromeria</i>	AJ704225	AJ704237	AJ704248
<i>Ditassa mucronata</i>	AJ704279	AJ704278	AJ704259
<i>Ditassa niruri</i>	AJ699340	AJ428752	AJ428751
<i>Ditassa retusa</i>	AJ704280	AJ704282	AJ704283
<i>Ditassa tomentosa</i>	AJ704483	AJ704486	AJ704484
<i>Gonioanthea hilariana</i>	AJ704488	AY163688*	
<i>Hemipogon acerosus</i>	AJ704288	AJ704290	AJ704291
<i>Hemipogon andinum</i>	AJ704295	AJ704294	AJ704292
<i>Hemipogon hatschbachii</i>	JN574498	JN574637*	
<i>Hemipogon sprucei</i>	AJ704296	AJ704298	AJ704299
<i>Metastelma fiebrigii</i>	HF547188	HF547134	
<i>Metastelma latifolium</i>	HF547190	HF547137*	
<i>Metastelma linearifolium</i>	AJ699341	AJ428809	AJ428808
<i>Metastelma oranense</i>	HF547197	HF547145*	
<i>Metastelma palmeri</i>	HF547199	HF547148*	
<i>Metastelma schaffneri</i>	AJ699343	AJ410216	AJ410215
<i>Metastelma schlechtendalii</i>	HF547207	HF547156*	
<i>Metastelma tubatum</i>	HF547216	HF547164*	
<i>Minaria acerosa</i>	AJ699285	AJ699287	AJ699288
<i>Minaria cordata</i>	AJ699300	AJ699299	AJ699297
<i>Minaria decussata</i>	DQ026707	AY163677*	
<i>Minaria grazielae</i>	AJ699339	AJ410204	AJ410203
<i>Nautonia nummularia</i>	AJ699344	AJ410228	AJ410227
<i>Nephradenia acerosa</i>	AJ704496	AY163705	AY163704
<i>Nephradenia asparagoides</i>	AJ704499	AY163707	AY163706
<i>Peplonia asteria</i>	AJ704303	AJ704302	AJ704300
<i>Petalostelma martianum</i>	JN574509	JN574648*	
<i>Petalostelma sarcostemma</i>	AJ699345	AJ428788	AJ428787
Tassadiinae			
<i>Tassadia berteriana</i>	AJ699336	AJ428791	AJ428790
<i>Tassadia guanchezii</i>	LN555752	LN555748*	
<i>Tassadia obovata</i>	AJ699284	AY163724	AY163723
<i>Tassadia richardiana</i>	HF547198	HF547146*	
Oxypetalinae			
<i>Araujia angustifolia</i>	AJ704333	AJ704332	AJ704330
<i>Araujia plumosa</i>	AJ704334	AJ704336	AJ704337
<i>Araujia sericifera</i>	AJ699352	AJ428794	AJ428793
<i>Funastrum angustifolium</i>	AJ699353	AJ428761	AJ428760
<i>Funastrum odoratum</i>	AJ699356	AJ290869	AJ290870
<i>Morrenia odorata</i>	AJ704342	AJ704344	AJ704345
<i>Oxypetalum coccineum</i>	AJ704327	AJ704326	AJ704329

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APPENDIX 1. (Continued)

Species	rps16	trnL intron	trnL-trnF spacer
<i>Oxypetalum coeruleum</i>	AJ704357	AJ704356	AJ704354
<i>Oxypetalum lanatum</i>	AJ704506	AJ704508	AJ704507
<i>Oxypetalum minarum</i>	AJ704511	AY163713*	
<i>Oxypetalum pannosum</i>	AJ704512	AJ704514	AJ704513
<i>Oxypetalum sublanatum</i>	AJ704517	AY163715*	
<i>Oxypetalum warmingii</i>	AJ704518	AJ704520	AJ704519
<i>Philibertia boliviana</i>	AJ704230	AJ704232	AJ704233
<i>Philibertia candolleana</i>	AJ699357	AJ410177	AJ410176
<i>Philibertia discolor</i>	AJ704526	AY163700*	
<i>Philibertia fontellae</i>	AJ699358	AJ492154	AJ492153
<i>Philibertia gilliesii</i>	AJ699359	AJ290899	AJ290898
<i>Philibertia globiflora</i>	AJ704238	AJ704236	AJ704234
<i>Philibertia lysimachioides</i>	DQ026714	AJ290900	AJ290901
<i>Philibertia multiflora</i>	AJ704246	AJ704245	AJ704243
<i>Philibertia parviflora</i>	AJ699361	AJ410225	AJ410224
<i>Philibertia vaileae</i>	AJ699362	AJ290905	AJ290904
<i>Tweedia brunonis</i>	AJ704256	AJ704258	AJ704260
Gonolobinae			
<i>Anemotrochus viridivenius</i>	LN901514	LN901496*	
<i>Anemotrochus yamanigueyensis</i>	LN901526	LN929899	LN929898
<i>Dictyanthus macvaughianus</i>	MH464890	MH464896*	
<i>Dictyanthus parviflorus</i>	EU038871	EU031617	
<i>Dictyanthus pavonii</i>	MH464892	MH464898*	
<i>Dictyanthus reticulatus</i>	EU038868	EU031614	
<i>Fischeria panamensis</i>	EU038832	EU031577	
<i>Fischeria scandens</i>	EU038833	EU031578	
<i>Gonolobus albomarginatus</i>	EU038834	EU031579	
<i>Gonolobus arizonicus</i>	EU038835	EU031580	
<i>Gonolobus barbatus</i>	AJ704264	AJ704263	AJ704261
<i>Gonolobus bibarbatus</i>	EU038836	EU031581	
<i>Gonolobus breedlovei</i>	EU038837	EU031582	
<i>Gonolobus chloranthus</i>	EU038838	EU031583	
<i>Gonolobus croceus</i>	EU038839	EU031584	
<i>Gonolobus fraternus</i>	EU038840	EU031585	
<i>Gonolobus fuscoviolaceus</i>	EU038841	EU031586	
<i>Gonolobus gonoloboides</i>	EU038842	EU031587	
<i>Gonolobus grandiflorus</i>	EU038843	EU031588	
<i>Gonolobus iyanolensis</i>	EU038844	EU031589	
<i>Gonolobus jaliscensis</i>	EU038845	EU031590	
<i>Gonolobus jamaicensis</i>	EU038846	EU031591	
<i>Gonolobus niger</i>	EU038847	EU031592	
<i>Gonolobus ophioglossa</i>	LN901515	LN901497*	
<i>Gonolobus parviflorus</i>	DQ026712	AY163689*	
<i>Gonolobus pectinatus</i>	EU038848	EU031593	
<i>Gonolobus rostratus</i>	EU038849	EU031594	

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APPENDIX 1. (Continued)

Species	<i>rps16</i>	<i>trnL</i> intron	<i>trnL-trnF</i> spacer
<i>Gonolobus stapelioides</i>	EU038850	EU031595	
<i>Gonolobus stenosepalus</i>	EU038851	EU031596	
<i>Gonolobus stephanotrichus</i>	EU038852	EU031597	
<i>Gonolobus suberosus</i>	EU038854	AJ704276	AJ704277
<i>Gonolobus uniflorus</i>	EU038855	EU031601	
<i>Gonolobus waitukubuliensis</i>	EU038856	EU031602	
<i>Gonolobus youroumaynensis</i>	EU038857	EU031603	
<i>Ibatia maritima</i>	EU038875	EU031621	
<i>Ibatia rubra</i>	EU038885	EU031632	
<i>Macroscepis diademata</i>	AJ704265	AJ704267	AJ704268
<i>Matelea acuminata</i>	EU038858	EU031604	
<i>Matelea angustiloba</i>	EU038859	EU031605	
<i>Matelea bicolor</i>	EU038860	LN901498*	
<i>Matelea biflora</i>	KF539850	KY582366	
<i>Matelea chrysantha</i>	EU038861	EU031607	
<i>Matelea correllii</i>	EU038864	EU031610	
<i>Matelea cyclophylla</i>	EU038865	AJ704272	AJ704269
<i>Matelea cynanchoides</i>	KY449041	KY582368	
<i>Matelea denticulata</i>	EU038867	EU031613	
<i>Matelea ekmanii</i>	EU038869	EU031615	
<i>Matelea inconspicua</i>	EU038872	EU031618	
<i>Matelea lanata</i>	EU038873	EU031619	
<i>Matelea lanceolata</i>	EU038874	EU031620	
<i>Matelea nipensis</i>	LN901517	LN901543	LN901550
<i>Matelea oblongata</i>	EU038877	EU031623	
<i>Matelea parviflora</i>	KY449038	KY582371	
<i>Matelea pauciflora</i>	EU038878	EU031625	
<i>Matelea pedalis</i>	AJ704530	AY163699*	
<i>Matelea phainops</i>	EU038879	EU031626	
<i>Matelea producta</i>	KY449036	KY582365	
<i>Matelea prosthecidiscus</i>	EU038880	EU031627	
<i>Matelea pubiflora</i>	EU038881	EU031628	
<i>Matelea pusilliflora</i>	EU038882	EU031629	
<i>Matelea reticulata</i>	EU038884	EU031631	
<i>Matelea sagittifolia</i>	KY449037	KY582372	
<i>Matelea sintenisii</i>	EU038886	EU031633	
<i>Matelea tamnifolia</i>	EU038887	LN901499*	
<i>Matelea texensis</i>	KY449039	KY582376	
<i>Matelea trachyantha</i>	EU038888	EU031635	
<i>Matelea variifolia</i>	EU038889	EU031636	
<i>Polystemma canisferum</i>	KY449035	KY582364	
<i>Polystemma cordatum</i>	EU038862	EU031608	
<i>Polystemma cordifolium</i>	EU038863	EU031609	
<i>Polystemma guatemalense</i>	KY449043	KY582353	
<i>Polystemma viridiflorum</i>	EU038866	EU031612	

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APPENDIX 1. (Continued)

Species	<i>rps16</i>	<i>trnL</i> intron	<i>trnL-trnF</i> spacer
<i>Schubertia grandiflora</i>	AJ699364	AJ428827	AJ428826
<i>Tylodontia fuscula</i>	LN901520	LN901544	LN901551
<i>Tylodontia urceolata</i>	LN901521	LN901545	LN901552
<i>Urostephanus gonoloboides</i>	EU038870	EU031616	
<i>Urostephanus patalensis</i>	MH464894		

APPENDIX 2. Information (taxon, collector, and coll. number (herbarium)) for specimens used in the distribution map.

Dictyanthus asper (Mill.) W.D. Stevens: J. Rzedowski 36889a (ENCB), J.C. Soto N. 982 (MEXU, MO, TEX), J.C. Soto N. 3491 (MEXU, MO); *Dictyanthus macvaughianus* (W.D. Stevens) W.D. Stevens: J.N. Labat 321 (P), J.N. Labat 1749 (P), R. McVaugh 24934 (MICH), E. Pérez C. & E. García L. 1359 (MEXU); *Dictyanthus parviflorus* Hemsl.: R.L. Oliver et al. 865 (MO), G. Arsene s.n. (G); *Dictyanthus pavonii* Decne.: C.P. Cowan 5686 (MO, TEX), H.G. Galeotti 1394 (MO), G.B. Hinton 13275 (LL, MO), G.B. Hinton 15061 (LL, MO), A.L. Mirón M. & G. Cornejo T. 369 (MEXU, MO), J.C. Soto N. et al. 9393 (MEXU).