# Amana wanzhensis (Liliaceae), a new species from Anhui, China 

BANGXING HAN ${ }^{1,2}$, KE ZHANG $^{3}$ \& LUQI HUANG ${ }^{2 *}$<br>${ }^{1}$ College of Biological and Pharmaceutical Engineering; Research Center of Research and Development of Traditional Chinese Medicine; West Anhui University; Anhui Province, Lu'an, 237012, China<br>${ }^{2}$ National Resource Center for Chinese Materia Medica, China Academy of Chinese Medical Science, State Key Laboratory of Dao-di Herbs, Beijing 100700, China<br>${ }^{3}$ Department of Pharmacy, Anhui University of Chinese Medicine, Hefei 230031, China; e-mail: huangluqi01@126.com<br>*Author for correspondence


#### Abstract

Amana wanzhensis, a new species from Ningguo County, Eastern China, is described and illustrated. A. wanzhensis is similar to $A$. erythronioides in sharing villous tunics and oblanceolate leaves, but differs from it by having shorter bracts $(0.1-0.5 \mathrm{~cm}$ long), yellow anthers, and deciduous tepals.


Keywords: ITS, Lilioideae, taxonomy, $\operatorname{trn} \mathrm{L}$ intron, Tulipeae

## Introduction

The genus Amana Honda (1935: 20) includes 4-5 species, endemic to Eastern Asia (Tan et al. 2007). This genus is overlapping with Tulipa Linnaeus (1753: 305) in many morphological character-states, and generic delimitation is confused. Many taxonomists considered Amana and Tulipa as synonyms (Sealy 1957, Mao 1980, Ohwi 1992, Tamura 1998, Shen 2001); however, other authors treated them as two distinct genera based on morphological characters and biogeography (Wu 2003), morphological (Tan et al. 2005) and molecular phylogenies (Peruzzi et al. 2009 and literature cited therein). Amana is characterized by $2-3(-4)$ opposite or verticillate bracts at the upper part of flowering stem, and is endemic to eastern Asia. Tulipa has no bracts on scape and distributed from Middle Asia to West Europe (Christenhusz et al. 2013).

There are four species of Amana in China (Wu 2003). Recently, one new species, Amana kuocangshanica D.Y.Tan \& D.Y.Hong in Tan et al. (2007: 443) has been discovered.

During our fieldwork in Ningguo County, Ahui Province, China, in 2012, a unknown species with a lot of populations was discovered. Our further examination and analysis indicated that it was a new species by having a unique combination of character-states in Amana.

## Materials and methods

Population sampling:-A total of 15 individuals of 4 Amana species were sampled, and at least two individuals were collected for each species (Table 1). All sequences from this study and two ITS region sequences (EU912095 and HE656028) from A. erythronioides (Baker) D.Y.Tan \& D.Y.Hong in Tan et al. (2007: 441) were used as ingroups, while two sequences (JQ776498 and JQ280387) from Tulipa saxatilis Baker (1883: 168) was used to be combined one sequence as outgroup for the phylogenetic analysis. New sequences for Amana were produced in the present study, while all other sequences were retrieved from GenBank. The specimens and GenBank accession number in this study are listed in Table 1.
TABLE 1. Specimens and GenBank accession number in this study.

| Taxon and samples | Collection site | Voucher number | Haplotype number | GenBank accession number (Reference) |  |
| :--- | :--- | :--- | :--- | :--- | :--- |
|  |  |  |  | trnL intron |  |
| A. wanzhensis | Ningguo, Anhui, China | Han B.X. N2 | Hap1 | KJ402416 (In the present study) | KJ402424 (In the present study) |
| A. wanzhensis | Ningguo, Anhui, China | Han B.X. N3 | Hap2 | KJ402417 (In the present study) | KJ402425 (In the present study) |
| A. wanzhensis | Ningguo, Anhui, China | Han B.X. N4 | Hap2 | KJ402418 (In the present study) | KJ402426 (In the present study) |
| A. wanzhensis | Ningguo, Anhui, China | Han B.X. N5 | Hap2 | KJ402419 (In the present study) | KJ402427 (In the present study) |
| A. edulis | Chuzhou, Anhui, China | Han B.X. C1 | Hap3 | KJ402420 (In the present study) | KJ402428 (In the present study) |
| A. edulis | Chuzhou, Anhui, China | Han B.X. C2 | Hap4 | KJ4402421 (In the present study) | KJ402429 (In the present study) |
| A. edulis | Chuzhou, Anhui, China | Han B.X. C3 | Hap5 | KJ402422 (In the present study) | KJ402430 (In the present study) |
| A. edulis | Chuzhou, Anhui, China | Han B.X. C5 | Hap6 | KJ402423 (In the present study) | KJ402431 (In the present study) |
| A. erythronioides | Huangshan, Anhui, China | Han B.X. F1 | Hap7 | EU912095 (From GenBank) | KJ402432 (In the present study) |
| A. erythronioides | Huangshan, Anhui, China | Han B.X. F2 | Hap8 | HE656028 (From GenBank) | KJ402433 (In the present study) |
| A. anhuiensis | Qianshan, Anhui, China | Han B.X. T1 | Hap9 | - | KJ402434 (In the present study) |
| A. anhuiensis | Qianshan, Anhui, China | Han B.X. T2 | Hap9 | - | KJ402435 (In the present study) |
| A. anhuiensis | Qianshan, Anhui, China | Han B.X. T3 | Hap9 | - | KJ402436 (In the present study) |
| A. anhuiensis | Qianshan, Anhui, China | Han B.X. T4 | Hap9 | - | KJ402437 (In the present study) |
| A. anhuiensis | Qianshan, Anhui, China | Han B.X. T5 | Hap9 | - | KJ402438 (In the present study) |
| Outgroup: Tulipa saxatilis | - |  | Hap10 | JQ776498 (From GenBank) | JQ280387 (From GenBank) |

DNA extraction, amplification, and sequencing:-Whole genomic DNA was extracted using the CTAB protocol from Rogers (1988). A partial nrDNA fragment of ITS region was amplified with the primers 17SE (5'ACGAATTCATGGTCCGGTGAAGTGTTC G-3') and 26SE (5'-TAGAATTCCCCGGTTCGCTCGCCGTTAC-3') (Sun et al. 1994; Clennett et al. 2012), while a partial cpDNA fragment of $t r n \mathrm{~L}$ intron was amplified with the primers c (5'-CGAAATCGGTAGACGCTACG-3') and d (5'-GGGGATAGAGGGACTTGAAC-3') (Taberlet et al. 1991). These DNA fragments were amplified using a standard polymerase chain raction (PCR), and then the purified amplified products were sequenced using both forward and reverse primers on an ABI-PRISMTM 310 Genetic Analyzer (Applied Biosystems Information, USA). Sequences were edited and aligned manually using BIOEDIT version 7.0.9.0 (Hall 1999).

Phylogenetic analysis:-Test for homogeneity of the nrDNA ITS region and cpDNA $\operatorname{trnL}$ intron data was performed using HomPart command in PAUP* version 4.0 beta 10 (Swofford 2002), and this test was described as the incongruence-length difference test (Farris et al. 1995). The homogeneity of nucleotide base frequencies across taxa was checked using the chi-square test implemented in PAUP* version 4.0 beta 10 (Swofford 2002). For phylogenetic analyses, Bayesian inference (BI) and maximum parsimony (MP) were performed in MrBayes version 3.1.2 (Ronquist \& Huelsenbeck 2003) and PAUP* version 4.0 beta 10 (Swofford 2002), respectively. For the Bayesian analysis, four partitions (i.e. partial ITS1, 5.8S, ITS2, and $\operatorname{trnL}$ intron sequences) were applied to the data, and models of molecular evolution were assessed for each partition using MrModeltest version 2.3 (Nylander 2004). The best-fit model (GTR) for partial ITS1, (K80) for 5.8 S , (HKY) for ITS2, and (HKY) for $t r n \mathrm{~L}$ intron were selected by the Akaike Information Criterion (AIC) in MrModeltest version 2.3 (Nylander 2004). Four Markov Chains Monte Carlo (MCMC) samples were run for $5 \times 106$ generations. Two independent runs were performed to allow additional confirmation of the convergence of MCMC runs. Trees were sampled every 100 generations, providing 105 samples from the two runs. Analysis of the standard deviation of split frequencies between the two runs was used to determine that stationarity had been reached after $5 \times 104$ generations, which were typically discarded as burn-in, leaving $9.9 \times 104$ samples to estimate the consensus tree and the Bayesian posterior probabilities. For the MP analysis, bootstrap analyses (Felsenstein 1988) were performed with 1000 replicates. Gaps were treated as missing data, and all characters had equal weight.

## Description of the new species

Amana wanzhensis L.Q.Huang, B.X.Han \& K.Zhang, sp. nov. (Fig. 1)

Haec species nova ad A. erythronioides affinis, sed bracteis minoribus (0.1-0.5 cm longis), antherae luteae, tepalis deciduis differt.

Type:-China. Anhui Province: Ningguo City, Xianxia Town, $30^{\circ} 34^{\prime} 79^{\prime \prime} \mathrm{N}, 119^{\circ} 22^{\prime} 97^{\prime \prime} \mathrm{E}$, alt. 735 m , 18 March 2013, B.X. Han \& X.W. Song 2012125 (holotype, ACM!, isotype, PE!).

Perennial herbs; bulbs ovoid, $1.5-2.5 \mathrm{~cm}$ in diameter, tunics brown, papery, pilose inside. Stems $15-30 \mathrm{~cm}$ tall, glabrous, simple. Leaves 2, opposite, lanceolate, green, $15-30 \mathrm{~cm}$ long, $1-3 \mathrm{~cm}$ wide, entire, obviously vein. Bracts usually 3 in number, not whorled, ribbon, $0.1-0.5 \mathrm{~cm}$ long, floral deciduous. Flowers solitary, funnel-shaped; tepals 6 , white, with a green blotch at the base and brown strips on the back; Stamens 6, two-wheeled, anthers $0.4-0.6 \mathrm{~cm}$ long, yellow, filaments $0.5-0.7 \mathrm{~cm}$ long, white. Oval ovaries, yellowish-green, 0.6 cm long, styles 1 cm long. Fruits triquetrous, $1-2 \mathrm{~cm}$ long, $0.5-1 \mathrm{~cm}$ wide. Flowers in February or March and fruits in March or April. The new species is closely related to $A$. erythronioides, but readily distinguished from it by having shorter bracts ( $0.1-0.5 \mathrm{~cm}$ long), yellow anthers, deciduous tepals.

Distribution and habitat:-A. wanzhensis is endemic to Xianxia Town, Ningguo City, Anhui Province, where it is widespread. It mostly grows in moist bamboo forests or meadow with elevation ranging from 600 to 800 m .


FIGURE 1. A. wanzhensis (A) plant, (B) tunics, (C) outer tepal, (D) inner tepal, (E) pedicel and ovary , (F) fruit. From holotype, drawn by Yun-Xi Zhu.

0.01 expected changes per site

FIGURE 2. The partitioned Bayesian phylogenetic tree based on the combination of partial ITS regions and trnL intron. Numbers above the branches represent Bayesian posterior probabilities (PP) and MP bootstrap (BS) values. Taxa are haplotypes; all haplotype designations are listed in Table 1, followed by the species and numbers of individuals from each species having that haplotype [e.g., A. anhuiensis (5)].

## Molecular phylogeny

Sequences, homogeneity between data partitions, and homogeneity of nucleotide base frequencies across taxa:-Twenty-three new sequences were obtained. Sequence lengths of partial ITS region and trnL intron were 617 bp and 497 bp , respectively. The combined alignment of partial ITS region and trnL intron was 1114 bp . Because the partition homogeneity test for the nrDNA ITS region and cpDNA $\operatorname{trnL}$ intron data showed character congruence ( $P=1.00$ ), we combined partial ITS region and $\operatorname{trn} \mathrm{L}$ intron to obtain 16 combined sequences (including outgroup taxa) revealing 10 haplotypes (Table 1) for the phylogenetic analyses. A chi-square test indicates that there was no significant compositional heterogeneity of bases among these haplotypes (chi-square $=46.81, d f=27, P>0.01$ ), and thus the biasing effects on the phylogenetic analyses could be eliminated (Jermiin et al. 2004).

Phylogenetic analyses:-The Bayesian and MP analyses based on the combination of partial ITS regions and trnL intron resulted in almost identical tree topologies, and the posterior probability (PP) values from the Bayesian analysis were all higher than the bootstrap (BS) values from the MP analysis. Figure 2 showed the partitioned Bayesian tree along with the PP and BS values obtained by MP methods. All haplotypes from A. erythronioides, A. wanzhensis, and A. edulis formed a single moderately supported clade ( $\mathrm{PP}=0.93, \mathrm{BS}=67$ ), while the haplotype corresponding to $A$. anhuiensis (Hap9) was found to be outgroup of this clade. All haplotypes from A. erythronioides formed a monophyletic group ( $\mathrm{PP}=0.99, \mathrm{BS}=92$ ), while all haplotypes from $A$. wanzhensis formed another monophyletic group ( $\mathrm{PP}=1.00$, $\mathrm{BS}=93$ ). A. erythronioides and $A$. wanzhensis were reciprocally monophyletic, and these two species formed a clade $(\mathrm{PP}=0.95, \mathrm{BS}=79)$ that was sister to A. edulis from which all haplotypes formed a monophyletic group $(\mathrm{PP}=1.00$, $\mathrm{BS}=95$ ). Phylogenetic relationships estimated using BI and MP methods were almost identical, and both methods suggested that $A$. wanzhensis is an independent lineage, distinct from $A$. wanzhensis.

## Acknowledgements

We thank Prof. De-Qun Wang for the help in the field work. We express our gratitude to Prof. Zhen-Yu Li, Xiao-Hua Jin for critical review of manuscript. The authors are also grateful to Yun-Xi Zhu for the drawings, Dr. Qun Zhao for phylogenetic analysis. This study was supported by the special fund for Traditional Chinese Medicine (NO. 20100700 $8 \backslash 201207002 \backslash 201407002$ ), special fund for public health of Traditional Chinese Medicine (NO. [2011]76\[2012]13\[2 $013] 135$ ) and special protection of biological diversity of department environmental protection of China.

## References

Christenhusz, M.J.M., Govaerts, R., David, J.C., Hall, T., Borland, K., Roberts, P.S, Tuomisto, A., Buerki, S., Chase, M.W. \& Fay, M.F. (2013) Tiptoe through the tulips - cultural history, molecular phylogenetics and classification of Tulipa (Liliaceae). Botanical Journal of the Linnean Society 172: 280-328.
Clennett, J.C.B., Chase, M.W., Forest, F., Maurin, O., \& Wilkin, P. (2012) Phylogenetic systematics of Erythronium (Liliaceae): morphological and molecular analyses. Botanical Journal of the Linnean Society 170: 504-528.
Felsenstein, J. (1988) Phylogenies from molecular sequences: inference and reliability. Annual Review of Genetics 22: 521-565.
Hall, T.A. (1999) BIOEDIT: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series 41: 95-98.
Jermiin, L.S., Ho, S.Y.W., Ababneh, F., Robinson, J., \& Larkum, A.W.D. (2004) The biasing effect of compositional heterogeneity on phylogenetic estimates may be underestimated. Systematic Biology 53: 638-643.
Mao, Z.M. (1980) Tulipa L. In: Wang, F.Z. \& Tang, J. (Eds.) Flora Reipublicae Popularis Sinicae 14. Science Press, Beijing, China, pp. 87-93.
Nylander, J.A.A. (2004) MRMODELTEST version 2.1. Computer program distributed by the author. Uppsala University, Uppsala.
Ohwi, J., \& Kitagawa, M. (1992) New Flora of Japan. Shibundo Co., Ltd. Publishers, Tokyo, 1716 pp.
Rogers, S.O., \& Bendich, A.J. (1988) Extraction of DNA from milligram amounts of fresh, herbarium and mummified plant tissues. Plant Molecular Biology 5: 69-76.
Ronquist, F., \& Huelsenbeck, J.P. (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics 19: 1572-1574.
Sealy, J.R. (1957) Tulipa edulis. Curtis's Botanical Magazine 171: 293.

Shen X.S. (2001) A new species of Tulipa (Liliaceae) from China. Acta Botanica Yunnanica 23: 39-40.
Sun, Y., Skinner, D.Z., Liang, G.H., \& Hulbert, S.H. (1994) Phylogenetic analysis of Sorghum and related taxa using internal transcribed spacers of nuclear ribosomal DNA. Theoretical and Applied Genetics 89: 26-32.
Swofford, D.L. (2002) PAUP*: Phylogenetic Analysis using Parsimony (* and Other Methods), version 4.0b 10. Sinauer, Sunderland.
Taberlet, P., Gielly, L., Pautou, G., \& Bouvet, J. (1991) Universal primers for amplication of three non-coding regions of chloroplast DNA. Plant Molecular Biology 17: 1105-1109.
Tamura, M.N. (1998) Liliaceae. In: Kubitzki K. (Eds.) The Families and Genera of Vascular Plants. Flowering Plants. -Monocotyledons Lilianae. Springer-Verlag, Berlin, Heidelberg, pp. 350-351.
Tan, D.Y. Zhang, Z., Li, X.R., \& Hong, D.Y. (2005) Restoration of the genus Amana Honda (Liliaceae) on the basis of cladistic analysis of morphological characters. Acta Phytotaxonomica Sinica 43: 262-270.
Tan, D.Y., Li, X.R., \& Hong, D.Y. (2007) Amana kuocangshanica (Liliaceae), a new species from south-east China. Botanical Journal of the Linnean Society 154: 435-442.
Wu, Z.Y., Lu, A.M., Tang, Y.C., Chen, Z.D., \& Li, D.Z. (2003) The Families and Genera of Angiosperms in China: a Comprehensive Analysis. Science Press, Beijing, 1209 pp.

